

Computing with Sequences and Ranges

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Sequences: representation

DNAMStringSet: Vector of sequences, e.g., sequence of each exon in the UCSC knownGene track

A *DNAMStringSet* instance of length 289969

width seq

```
[1]    354 CTTGCCGTCAGCCTTT...TCACAACCTAGGCCA
[2]    127 GTCCTGTCTCCCCC...CCCAGTGTTGCAGAG
[3]    109 GTGTGTGGTGATGCCA...CCCAGTGTTGCAGAG
...    ...
[289968] 109 GTGTGTGGTGATGCCA...CCCAGTGTTGCAGAG
[289969] 354 CTTGCCGTCAGCCTTT...TGACAACCTAGGCCA
```

- ▶ Acts like a *vector*, e.g., `length()`, `[`, `[[`
- ▶ Many methods – `methods(class="DNAMStringSet")` – e.g., `reverseComplement()`, `letterFrequency()`, ...

Sequences: packages

Biostrings General purpose biological sequence representation.

BSgenome Whole-genome representation.

ShortRead High-throughput sequencing.

Sequences: classes

- `DNASTring` Single DNA sequence, e.g., chromosome
- `DNASTringSet` Vector of DNA sequences. Actually, `XString`,
`XStringSet`: X could be DNA, RNA, AA)
- `BSgenome` Collection of (large) DNA sequences
- `ShortReadQ` High-throughput reads & their qualities

Sequences: file references

TwoBitFile, FaFile .2bit (in *rtracklayer*) or .fa (in *Rsamtools*)
indexed genome-scale fasta files.

FastqFile , e.g., *FastqStreamer* (in *ShortRead*)

Effectively manage large data

- ▶ *Restrict* input to specific genomic locations
- ▶ *Iterate* through large files in chunks

Sequences: annotations

*BSgenome.** packages

- ▶ E.g., *BSgenome.Hsapiens.UCSC.hg19*
- ▶ Packages containing whole-genome sequences for model organisms

AnnotationHub resources

- ▶ e.g., Ensembl FASTA files

Ranges: *GRanges* representation

```
> gr = exons(TxDb.Hsapiens.UCSC.hg19.knownGene); gr
```

```
GRanges with 289969 ranges and 1 metadata column:
```

	seqnames	ranges	strand	exon_id
	<Rle>	<IRanges>	<Rle>	<integer>
[1]	chr1	[11874, 12227]	+	1
[2]	chr1	[12595, 12721]	+	2
[3]	chr1	[12613, 12721]	+	3
...
[289967]	chrY	[59358329, 59359508]	-	277748
[289968]	chrY	[59360007, 59360115]	-	277749
[289969]	chrY	[59360501, 59360854]	-	277750

```
---  
seqinfo: 93 sequences (1 circular) from hg19 genome
```

GRanges

```
length(gr); gr[1:5]  
seqnames(gr)  
start(gr)  
end(gr)  
width(gr)  
strand(gr)
```

DataFrame

```
mcols(gr)  
gr$exon_id
```

Seqinfo

```
seqlevels(gr)  
seqlengths(gr)  
genome(gr)
```

- ▶ Data: aligned reads, called peaks, SNP locations, CNVs, ...
- ▶ Annotation: gene models, variants, regulatory regions, ...

Ranges: *GRangesList* representation

```
> gr1 = exonsBy(TxDb.Hsapiens.UCSC.hg19.knownGene, "tx", use.names=TRUE); gr1
```

```
GRangesList of length 82960:
```

```
$uc001aaa.3
```

```
GRanges with 3 ranges and 3 metadata columns:
```

	seqnames	ranges	strand	exon_id	exon_name	exon_rank
	<Rle>	<IRanges>	<Rle>	<integer>	<character>	<integer>
[1]	chr1	[11874, 12227]	+	1	<NA>	1
[2]	chr1	[12613, 12721]	+	3	<NA>	2
[3]	chr1	[13221, 14409]	+	5	<NA>	3

```
GRangesList  
(list of GRanges)  
length(gr1)  
gr1[1:3]  
shift(gr1, 1)  
range(gr1)
```

```
$uc010nxq.1
```

```
GRanges with 3 ranges and 3 metadata columns:
```

	seqnames	ranges	strand	exon_id	exon_name	exon_rank
[1]	chr1	[11874, 12227]	+	1	<NA>	1
[2]	chr1	[12595, 12721]	+	2	<NA>	2
[3]	chr1	[13403, 14409]	+	6	<NA>	3

```
GRanges  
gr1[[2]]  
gr1[["uc010nxq.1"]]
```

```
$uc010nxr.1
```

```
GRanges with 3 ranges and 3 metadata columns:
```

	seqnames	ranges	strand	exon_id	exon_name	exon_rank
[1]	chr1	[11874, 12227]	+	1	<NA>	1
[2]	chr1	[12646, 12697]	+	4	<NA>	2
[3]	chr1	[13221, 14409]	+	5	<NA>	3

Two kinds of fun!

```
introns =  
  psetdiff(range(gr1), gr1)
```

```
grr = unlist(gr1)  
## transform grr, then...  
gr1 = relist(grr, gr1)
```

‘flesh’ ‘skeleton’

```
...  
<82957 more elements>
```

```
----
```

```
seqinfo: 93 sequences (1 circular) from hg19 genome
```


Ranges: packages

GenomicRanges Essential representation and operations

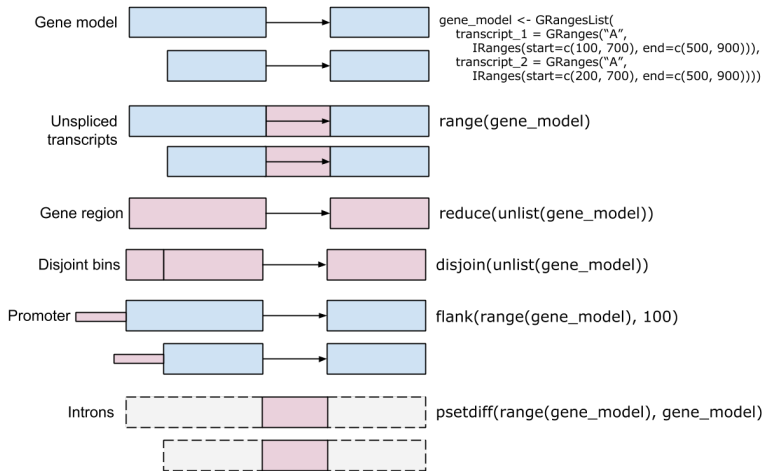
GenomicAlignments Aligned reads as genomic ranges

GenomicFeatures Annotations as genomic ranges

rtracklayer Annotation (e.g., BED, GTF) input

A little more advanced usage: *IRanges* (); *S4Vectors* (underling conceptual ideas)

Ranges: operations



- ▶ Many more, e.g., `methods(class="GRanges")`

Ranges: `findOverlaps()`

- ▶ Overlaps between query and subject genomic ranges
- ▶ Different types of overlap, e.g., 'any', 'within', ...

```
> q <- GRanges("chr1", IRanges(10, 20))
> s <- GRanges("chr1", IRanges(5, width=c(3, 6, 9)))
> findOverlaps(q, s)
```

Hits object with 2 hits and 0 metadata columns:

```
      queryHits subjectHits
      <integer>  <integer>
[1]           1           2
[2]           1           3
```

```
queryLength: 1
subjectLength: 3
```

- ▶ *Hits* object describing many-to-many relationship between overlapping ranges.

Ranges: working with files

`import` (*rtracklayer*) for BED, GTF, and other common web file import functions. *BEDFile*, *GTFFile*, etc.

`readGAlignments / readGAlignmentsList` (*GenomicAlignments*) for aligned reads in BAM files

`BamFile` (*Rsamtools*) for lower-level access to BAM files, e.g., restriction and iteration

Ranges: annotation

*TxDb.** packages

- ▶ E.g., *TxDb.Hsapiens.UCSC.hg19.knownGene*
- ▶ Genomic ranges for exons, transcripts, coding sequences, and how these are ordered into gene models, e.g., exons grouped by transcript

AnnotationHub resources

- ▶ Ensembl gene models
- ▶ Roadmap Epigenomics regulatory marks
- ▶ Many other range-based resources

What's to love about a data.frame?

- ▶ Coordinated data management
- ▶ Familiarity
- ▶ Interoperability

And yet...

- ▶ Not all columns are equal – seqlevels, start, width, strand are required, other columns are optional.
- ▶ Columns have special meaning – e.g., widths of genomic ranges are ≥ 0
- ▶ Not every column is a base R vector – *DNAStrngSet*
- ▶ Data from different tables are often related – information about genomes

Principled versus precocious?

- ▶ The tidyverse as principled – a few well-designed orthogonal functions operating exclusively (?) on `data.frame` derivatives
- ▶ *R* and *Bioconductor* as precocious – a large number of classes and specialized functions
- ▶ Some of precociousness is lack of principle, but some reflects high-level summary of complex work flows

Other resources

- ▶ [Workflows](#) & package vignettes
- ▶ [GenomicRanges](#) and other 'cheat sheets'
- ▶ Course material
- ▶ Support site [tutorials](#)

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