

Genotype and DNA Copy Number Estimation

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Very large data sets

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Statistical Methods for Very Large Datasets Conference 2011

Wednesday, June 01, 2011 8:00 AM -
Friday, June 03, 2011 5:30 PM (Eastern Time)

InterContinental Harbor Court Hotel
800-824-0076
550 Light Street
Baltimore, Maryland 21202
 [Map and Directions](#)

Welcome!



The Department of Biostatistics at the Johns Hopkins Bloomberg School of Public Health invites you to a 3-day conference on Very Large Data Sets. The conference is scheduled from June 1-3, 2011 and will be hosted in beautiful, downtown [Baltimore, Maryland](#), USA at the [InterContinental Harbor Court Hotel](#).

The conference has a one-track session for invited presentations and a high profile session for contributed poster presentations. A panel discussion will attempt to define what large data sets are, anticipate new challenges, and identify possible solutions.


Conference Overview


There is an acute and increasing need to adapt standard statistical methods and to develop new approaches for the analysis of very large data sets. A data set is very large if it raises difficult or insurmountable computational problems for standard data analysis using available computing systems. The continuous increase in size and complexity of data sets is due in part to increased computational and storage capabilities, new measurement technologies and study designs, and an increasing number of study "units".

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 77 days left to take advantage of early price.

Contact Information

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Important Dates:

June 1, 2010: Call for poster presentation abstracts
February 1, 2011: Final date for a early bird registration fee
March 1, 2011: Final date for submission of poster abstracts
April 1, 2011 : Notification for poster abstract acceptance
May 3, 2011: Final date for reduced conference rate at hotel
June 1-3, 2011: Conference dates

Confirmed Speakers include:

Goncalo Abecasis, *University of Michigan*
DuBois Bowman, *Emory University*
Brian Caffo, *Johns Hopkins University*
Raymond Carroll, *Texas A&M University*
Ciprian Crainiceanu, *Johns Hopkins University*
Francesca Dominici, *Harvard University*
William DuMouchel, *Phase Forward Lincoln Safety Group*
Sandrine Dudoit, *University of California at Berkeley*
Jay Emerson, *Yale University*
Stephen Eubank, *Virginia Tech*
Montse Fuentes, *North Carolina State University*
Robert Gentleman, *Fred Hutchinson Cancer Research Center*
Rafael Irizarry, *Johns Hopkins University*
Hongkai Ji, *Johns Hopkins University*
Nicole Lazar, *University of Illinois at Chicago*
Jeffrey Morris, *MD Anderson Cancer Center*
Hans-Georg Muller, *University of California at Davis*
Doug Nychka, *National Center for Atmospheric Research*
Todd Ogdén, *Columbia University*
Roger Peng, *Johns Hopkins University*
James Ramsay, *McGill University*
Ingo Ruczinski, *Johns Hopkins University*
Steven Salzberg, *University of Maryland*
Terry Speed, *University of California at Berkeley*
John Storey, *Princeton University*
Alex Szalay, *Johns Hopkins University*
Jonathan Taylor, *Stanford University*
Chris Volinsky, *AT&T Labs-Research*

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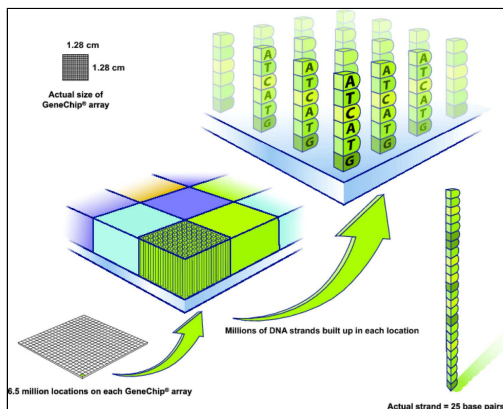
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Genomic arrays



Affymetrix SNP chip terminology

Genomic DNA: TACATAGCCATCGGTAN^AGTACTACTCAATGATGATA
SNP
G

PM probe for Allele A: ATCGGTAGCCAT^TCATGAGTTACTA

PM probe for Allele B: ATCGGTAGCCAT^CCATGAGTTACTA

Genotyping: answering the question about the two copies of the chromosome on which the SNP is located:

Is a person **AA**, **AG** or **GG** at this Single Nucleotide Polymorphism?

<http://www.affymetrix.com>

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Copy number estimates are noisy

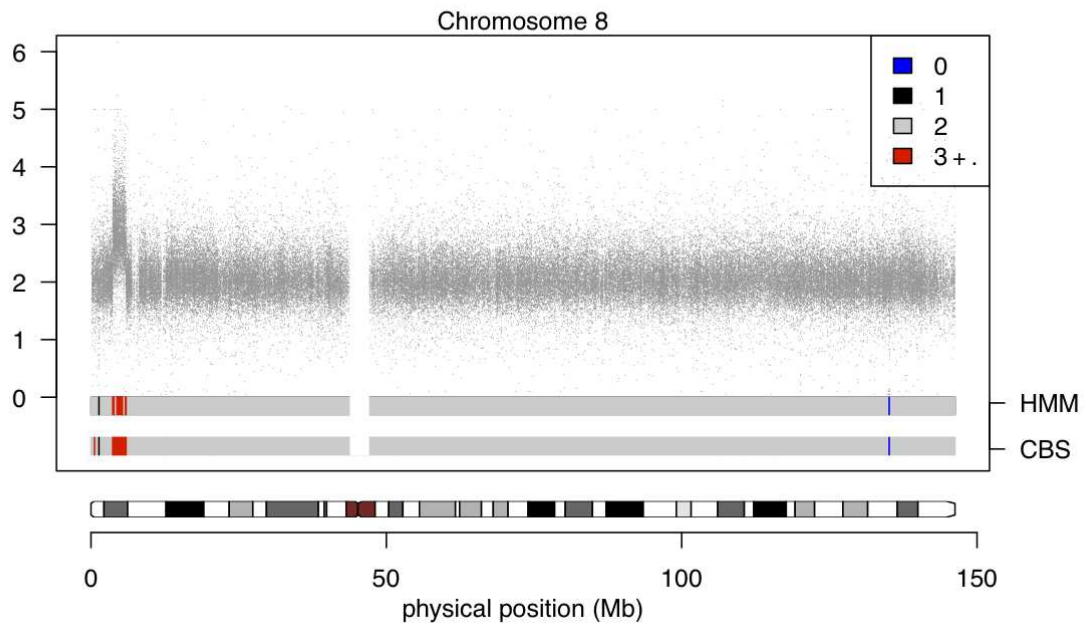
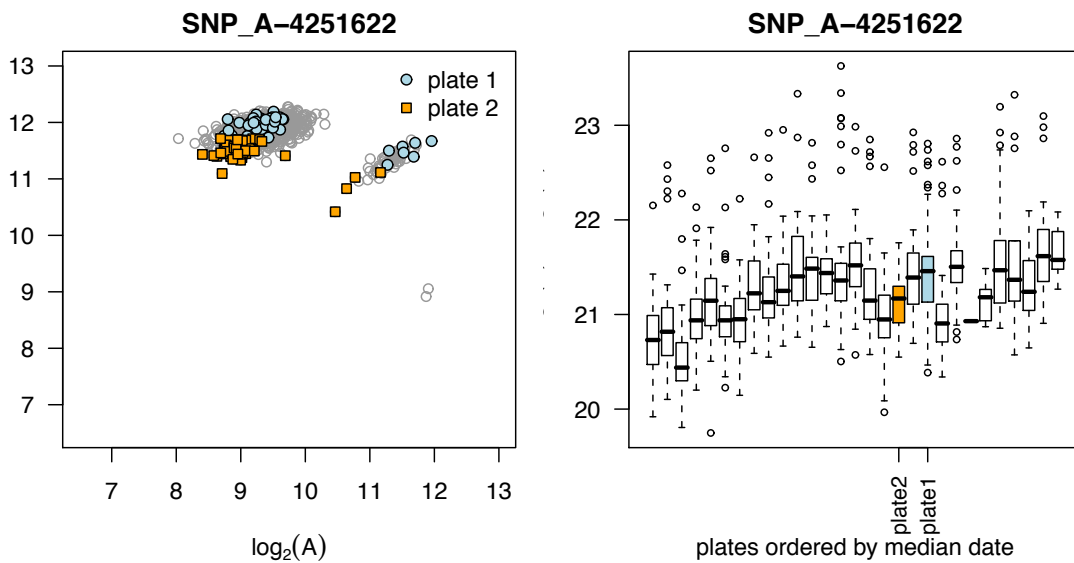
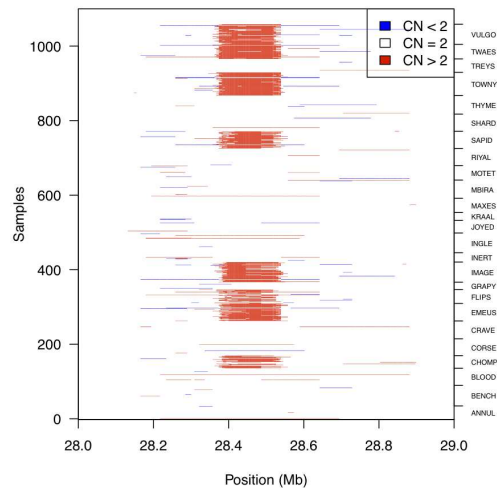
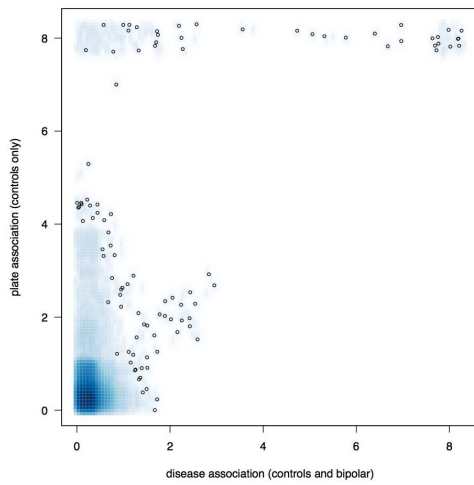


Plate effects



Confounding of plate and disease

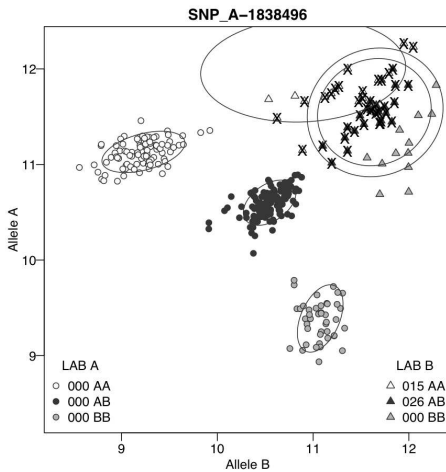


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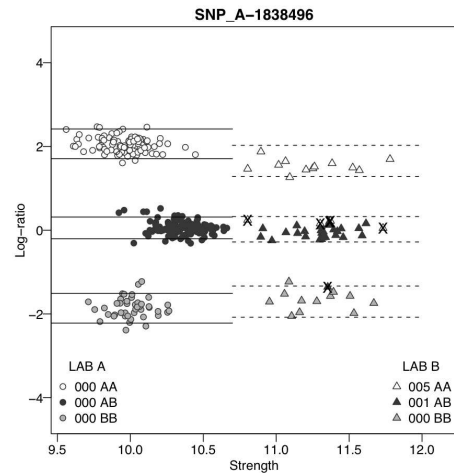
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Genotype estimates are more robust

Birdseed



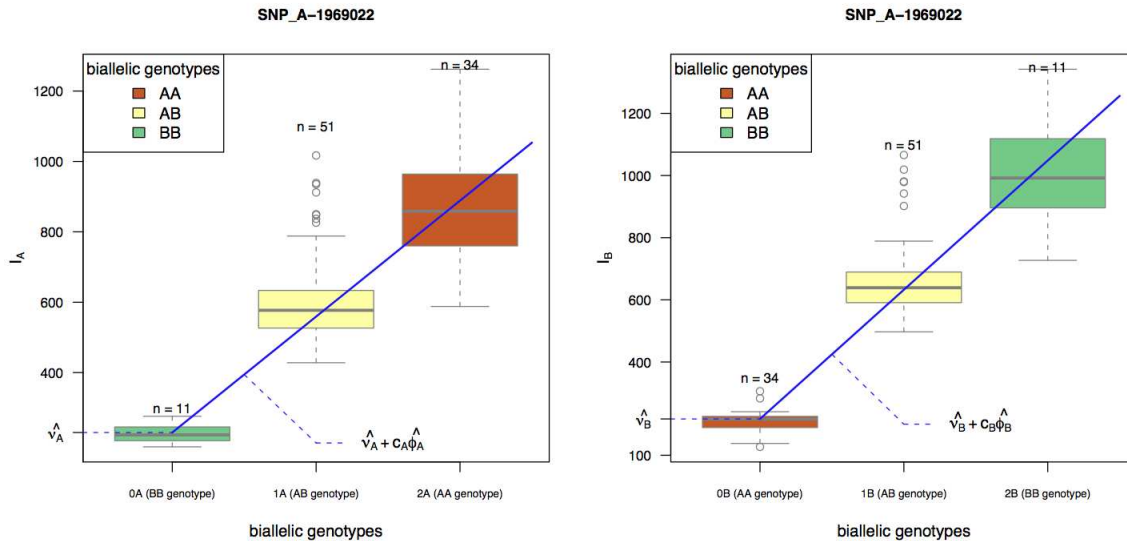
CRLMM



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Allele specific copy numbers



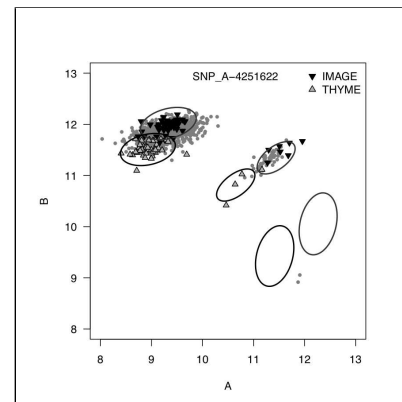
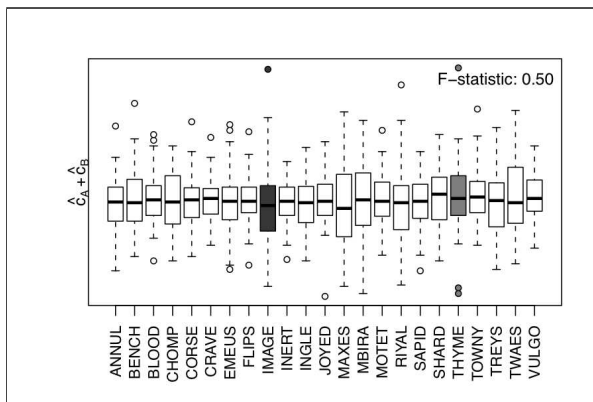
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Allele specific copy numbers

At locus i , for subject j in plate p , we have for allele $k \in \{A, B\}$

$$I_{kijp} = \nu_{kip} \delta_{kijp} + \phi_{kip} c_{kijp} \epsilon_{kijp} \implies \hat{c}_{kijp} = \max \left\{ \frac{1}{\hat{\phi}_{kip}} (I_{kijp} - \hat{\nu}_{kip}), 0 \right\}$$

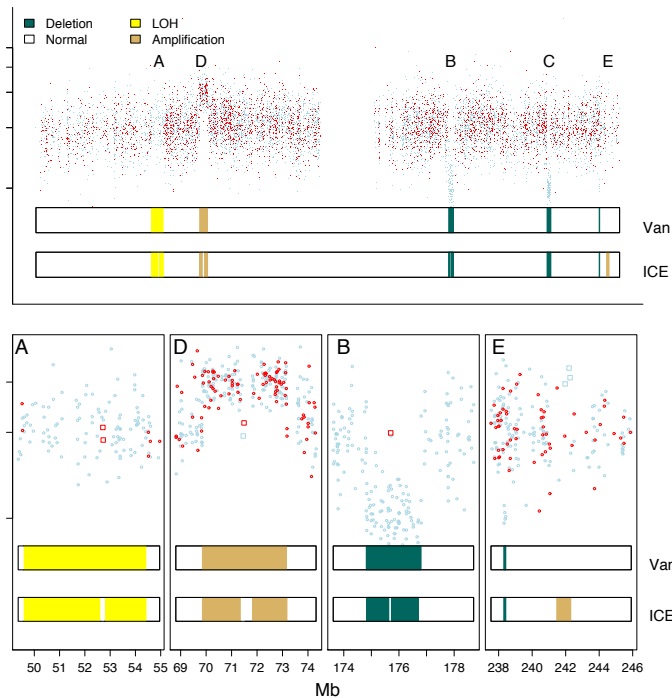


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Vanilla and ICE HMMs for genotype and copy number

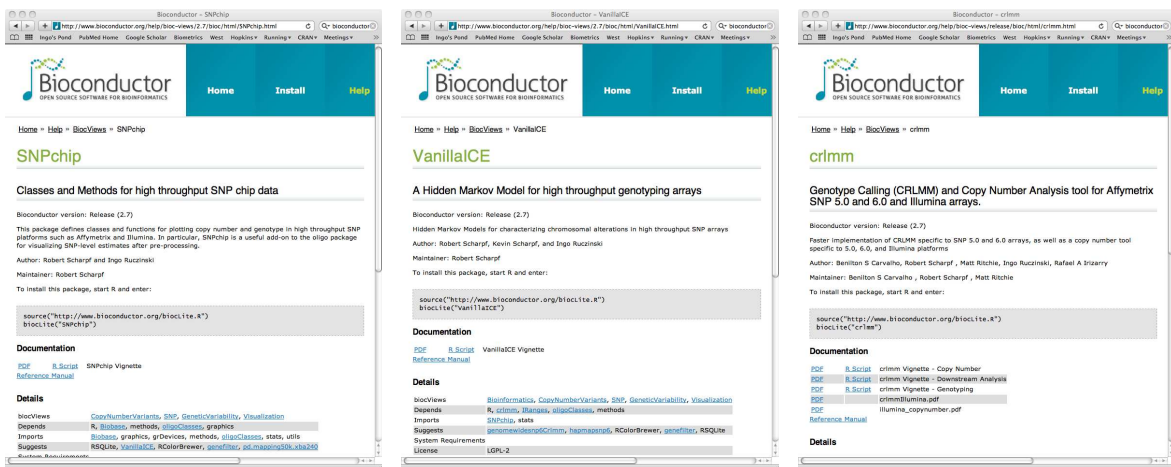


[SCH · PAR · PEV · RUC | AOAS 2008]

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Open source software



[SCH · · · RUC | BIOINF 2007] • [SCH · RUC | M·MOL·BIO 2010] • [SCH · RUC · · · IRI | BIostat 2010]

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Genotype and DNA copy number estimation

Using the R Package `crimm` for Genotyping and Copy Number Estimation

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Abstract

Genotyping platforms such as Affymetrix can be used to assess genotype-phenotype as well as copy number-phenotype associations at millions of markers. While genotyping algorithms are largely concordant when assessed on HapMap samples, tools to assess copy number changes are more variable and often discordant. One explanation for the discordance is that copy number estimates are susceptible to systematic differences between groups of samples that were processed at different times or by different labs. Analysis algorithms that do not adjust for batch effects are prone to spurious measures of association. The R package `crimm` implements a multilevel model that adjusts for batch effects and provides allele-specific estimates of copy number. This paper illustrates a workflow for the estimation of allele-specific copy number, develops marker- and study-level summaries of batch effects, and demonstrates how the marker-level estimates can be integrated with complimentary Bioconductor software for inferring regions of copy number gain or loss. All analyses are performed in the statistical environment R. A compendium for reproducing the analysis is available from the author's website (<http://www.biostat.jhsph.edu/~rscharpf/crimmCompendium/index.html>).

Keywords: copy number, batch effects, robust, multilevel model, high-throughput, oligonucleotide array.

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Compendium

Compendium for "Using the R Package `crimm` for Genotyping and Copy Number Estimation" by Scharpf, et al. (2010)

2.1 Reproducing the Figures

The `crimmCompendium` package contains the text, data, and R functions used to make the figures in this paper. Users should be able to reproduce the figures upon successful installation of the compendium. The compendium requires R ≥ 2.12 . To install the compendium and its dependencies you will need an internet connection.

```
source("http://www.bioconductor.org/biocLite.R")
pkgs <- c("crimm", "DNAcopy", "SNPchip", "RColorBrewer", "VanillaICE")
biocLite(pkgs)
install.packages("crimmCompendium_1.0.4.tar.gz", repos=NULL)
```

To install the `crimmCompendium`, download the tarball of the latest build:

R package	build
<code>crimmCompendium</code>	1.0.4

The package can be installed from the command line by R CMD `INSTALL crimmCompendium_1.0.4.tar.gz`, or from an R session in the same directory by:

```
install.packages("crimmCompendium_1.0.4.tar.gz", repos=NULL)
```

Windows users would first need to install the appropriate `Rpackage{Rtools}` executable.

R code extracted from the manuscript.Rnw vignette for reproducing the figures is available from the Code links adjacent to the figures below. To reproduce the figures, simply copy the code into R.

2.2 Reproducing the Manuscript

The complete analysis of the HapMap phase III data is contained in the manuscript.Rnw Sweave file. This document is located in the `inst/scripts` subdirectory of the `crimmCompendium` package. Three additional steps are required for the complete analysis. First, one must download and install the [HapMap Phase 3 CEL files](#) for the Affymetrix 6.0 platform. Secondly, one must change the following lines in the manuscript vignette as appropriate:

```
pathToCels <- "/your/path/to/CEL/Files"
outdir <- "/directory/to/store/results"
```

Finally, one must install additional package dependencies that were not required for installing the `crimmCompendium`. In particular, the packages `ff`, `genefilter`, `ellipse`, and `MASS`. Note that the genotyping and copy number estimation steps in the manuscript.Rnw Sweave file involve long computations. We suggest submitting the code using R CMD `batch`. Provided that LaTeX is installed, a pdf version of the manuscript can be generated by issuing the following commands from R:

```
library(tools)
texi2dvi("manuscript.tex", pdf=TRUE)
```

3 Figures and Code

Figures	R code
	Code

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