

# Introduction to DNA microarray technologies

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Bioconductor short course  
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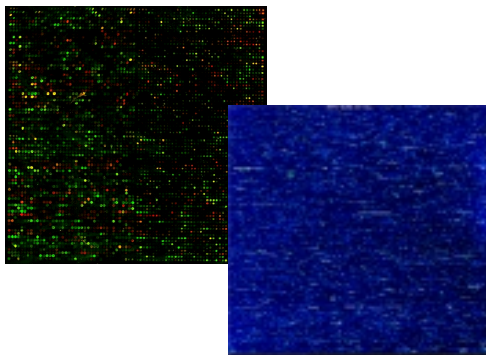
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## Outline

- Basic principles
- cDNA microarrays
- Affymetrix oligonucleotide chips

## DNA microarrays



## DNA microarrays

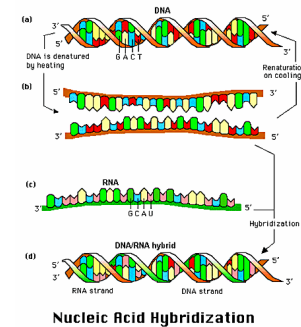
**DNA microarrays** rely on the **hybridization** properties of nucleic acids to monitor DNA or RNA abundance on a genomic scale in different types of cells.

The ancestor of cDNA microarrays: the **Northern blot**.

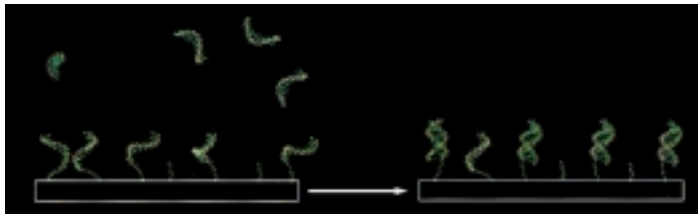
## Hybridization

- **Hybridization** refers to the **annealing** of two nucleic acid strands following the base pairing rules.
- Nucleic acid strands in a duplex can be separated, or **denatured**, by heating to destroy the hydrogen bonds.

## Hybridization



## Hybridization



## Gene expression assays

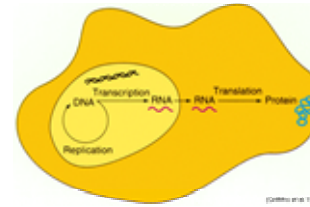
The main types of gene expression assays:

- Serial analysis of gene expression (SAGE);
- **Short oligonucleotide arrays (Affymetrix);**
- Long oligonucleotide arrays (Agilent Inkjet);
- Fibre optic arrays (Illumina);
- **cDNA arrays (Brown/Botstein).**

## Applications of microarrays

- **Measuring transcript abundance (cDNA arrays);**
- Genotyping;
- Estimating DNA copy number (CGH);
- Determining identity by descent (GMS);
- Measuring mRNA decay rates;
- Identifying protein binding sites;
- Determining sub-cellular localization of gene products;
- ...

## Transcriptome



- mRNA or transcript levels sensitively reflect the state of a cell.
- Measuring protein levels (translation) would be more direct but more difficult.

## Transcriptome

- The **transcriptome** reflects
  - Tissue source: cell type, organ.
  - Tissue activity and state:
    - Stage of development, growth, death.
    - Cell cycle.
    - Disease vs. healthy.
    - Response to therapy, stress.

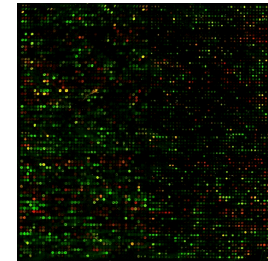
## Applications of microarrays

- **Cancer research:** Molecular characterization of tumors on a genomic scale
  - more reliable diagnosis and effective treatment of cancer.
- **Immunology:** Study of host genomic responses to bacterial infections; reversing immunity.
- ...

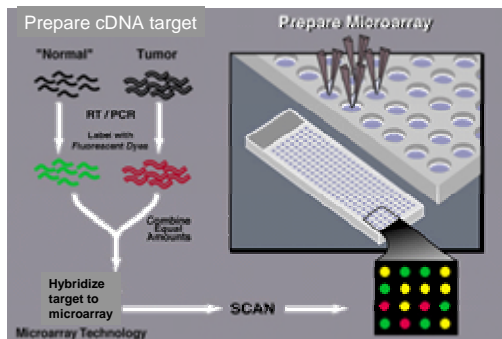
## Applications of microarrays

- Compare mRNA (transcript) levels in different types of cells, i.e., vary
  - Tissue: liver vs. brain;
  - Treatment: drugs A, B, and C;
  - State: tumor vs. non-tumor, development;
  - Organism: different yeast strains;
  - Timepoint;
  - etc.

## cDNA microarrays



## cDNA microarrays



## cDNA microarrays

- The **relative abundance** of a spotted DNA sequence in two DNA or RNA samples may be assessed by monitoring the **differential hybridization** of these two samples to the sequence on the array.
- **Probes**: DNA sequences spotted on the array, immobile substrate.
- **Targets**: Nucleic acid samples hybridized to the array, mobile substrate.

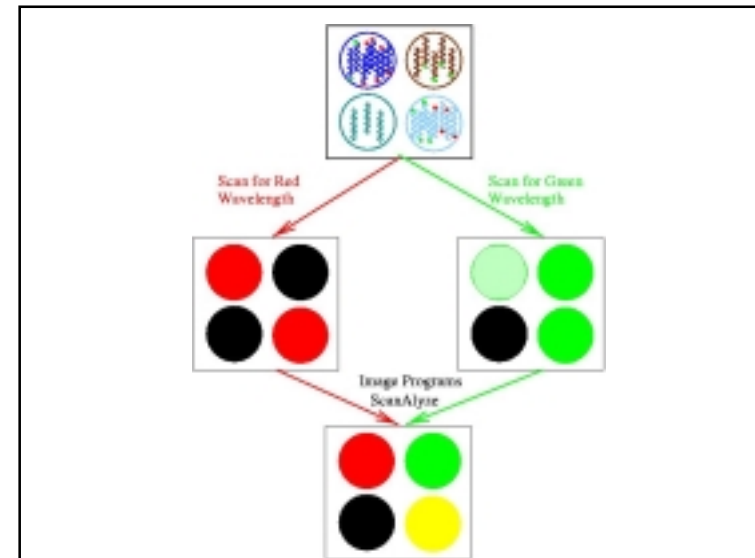
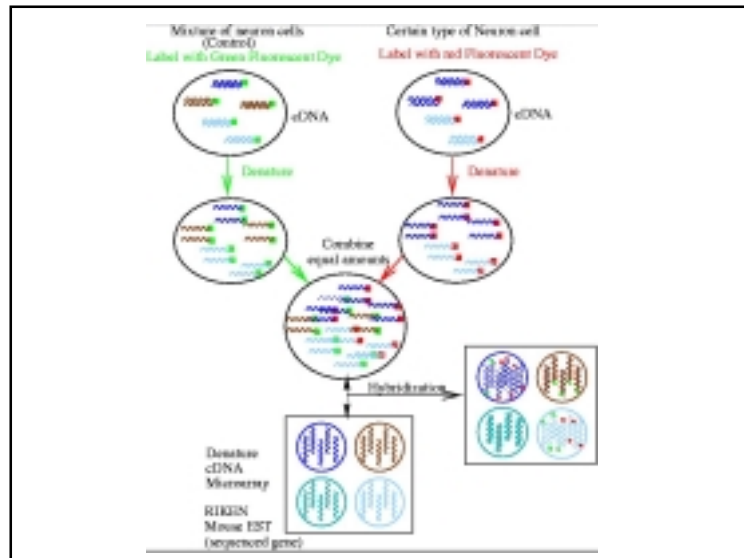
## cDNA microarrays

- The **ratio** of the red and green fluorescence intensities for each spot is indicative of the relative abundance of the corresponding DNA probe in the two nucleic acid target samples.

## cDNA microarrays

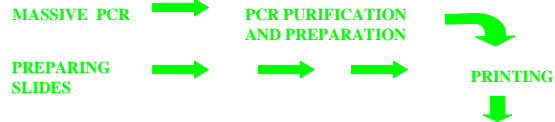
$$M = \log_2 R/G = \log_2 R - \log_2 G$$

- M < 0**, gene is over-expressed in green-labeled sample compared to red-labeled sample.
- M = 0**, gene is equally expressed in both samples.
- M > 0**, gene is over-expressed in red-labeled sample compared to green-labeled sample.

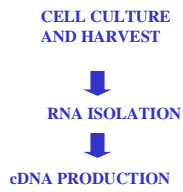


## The process

### Building the microarray:



### RNA preparation:



### Hybing the array:

#### ARRAY HYBRIDIZATION AND SCANNING

#### TARGET LABELING

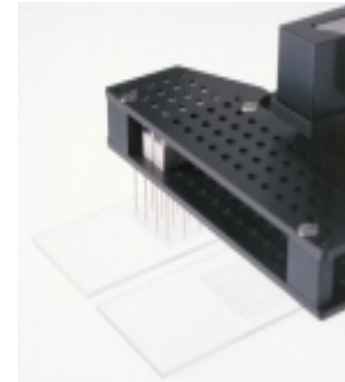
#### POST PROCESSING

#### DATA ANALYSIS

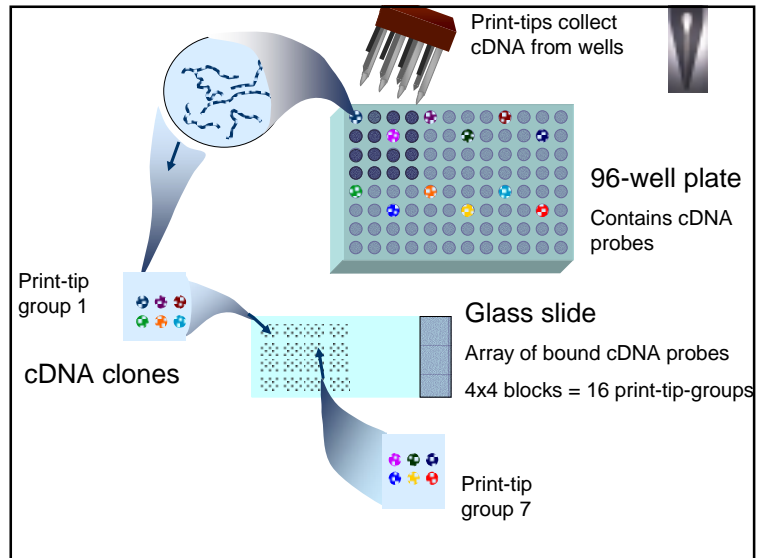
## The arrayer



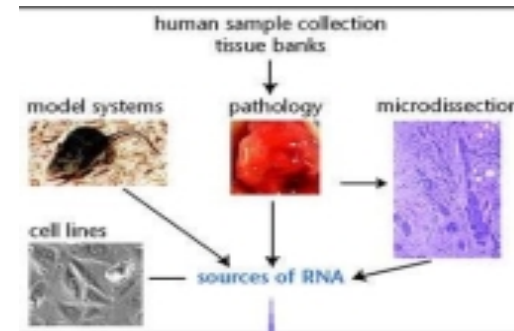
Ngai Lab arrayer, UC Berkeley



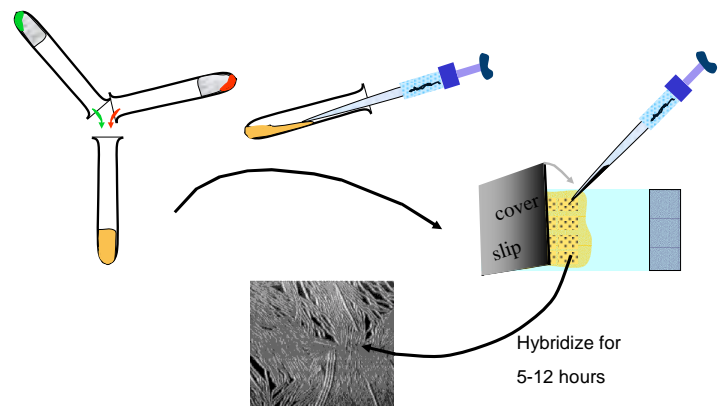
Print-head



## Sample preparation

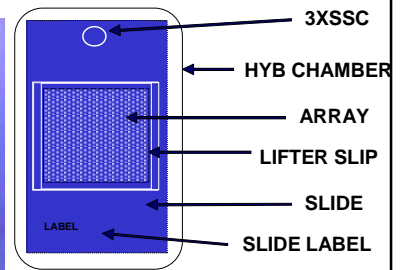


## Hybridization



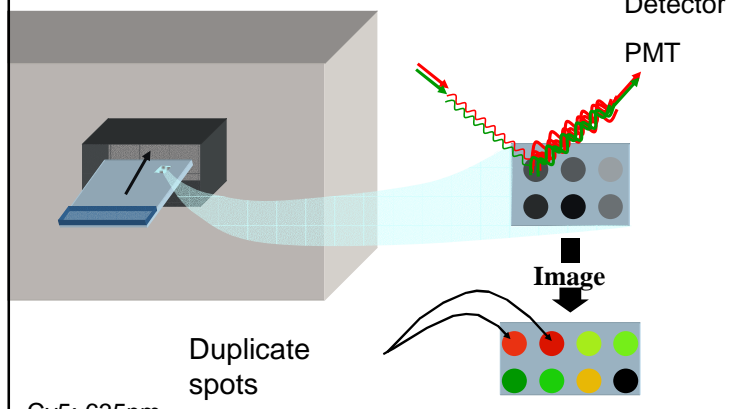
Binding of cDNA target samples to cDNA probes on the slide

## Hybridization chamber

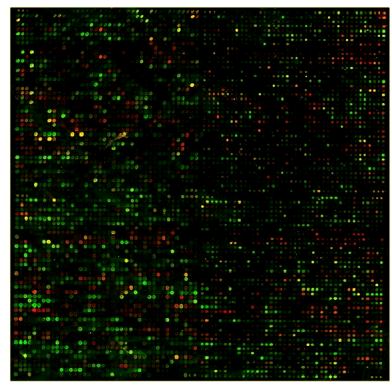


- Humidity
- Temperature
- Formamide (Lowers the Tmp)

## Scanning



## RGB overlay of Cy3 and Cy5 images



## Raw data

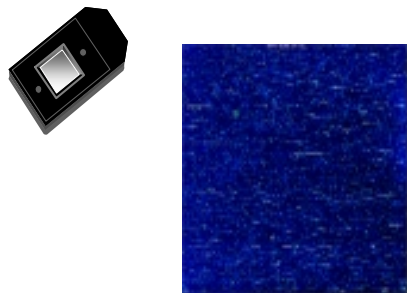
E.g. Human cDNA arrays

- ~43K spots;
- 16-bit TIFFs: ~ 20Mb per channel;
- ~ 2,000 x 5,500 pixels per image;
- Spot separation: ~ 136 $\mu$ m;
- For a “typical” array, the spot area has
  - mean = 43 pixels,
  - med = 32 pixels,
  - SD = 26 pixels.

## Animation

<http://www.bio.davidson.edu/courses/genomics/chip/chip.html>

## Oligonucleotide chips

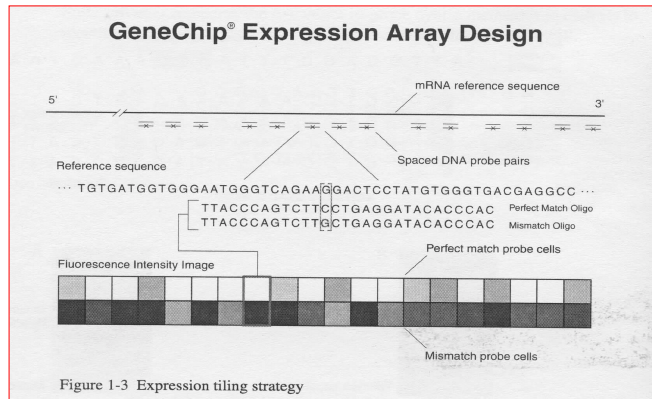


## Probe sets

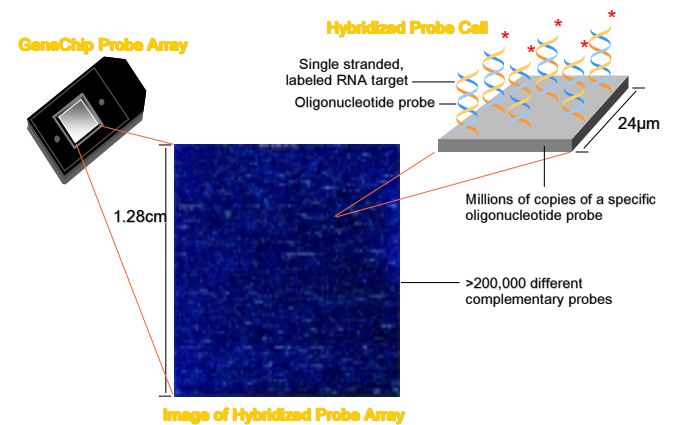
- Each gene is represented by 16-20 oligonucleotides of 25 base-pairs, i.e., 25-mers.
- **Perfect match probe, PM:** A 25-mer complementary to the reference sequence.
- **Mismatch probe, MM:** same as PM but with a single homomeric base change for the middle (13<sup>th</sup>) base.
- **Probe pair.** A (PM,MM) pair.
- **Probe set.** 16-20 probe pairs.
- The purpose of the MM probe design is to measure non-specific binding and background noise.



## Probe sets



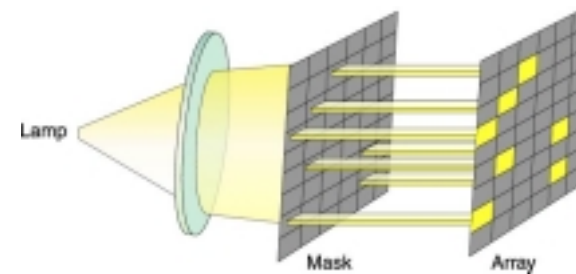
## Oligonucleotide chips



## Oligonucleotide chips

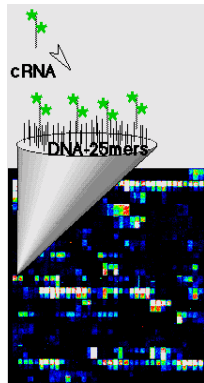
- The probes are synthesized *in situ*, using combinatorial chemistry and photolithography.
- **Probe cells** are square shaped features on the chip containing millions of copies of a single 25 ~~nt~~ probe. Sides are 18 ~~µ~~microns.

## Oligonucleotide chips



The manufacturing of GeneChip® probe arrays is a combination of photolithography and combinatorial chemistry.

## Image analysis

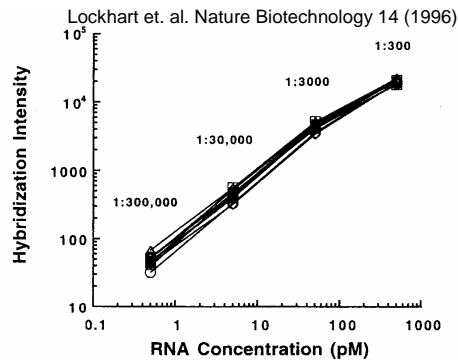


- About 100 pixels per probe cell.
- These intensities are combined to form one number representing the expression level for the probe cell oligo.
- → CEL file with PM or MM intensity for each cell.

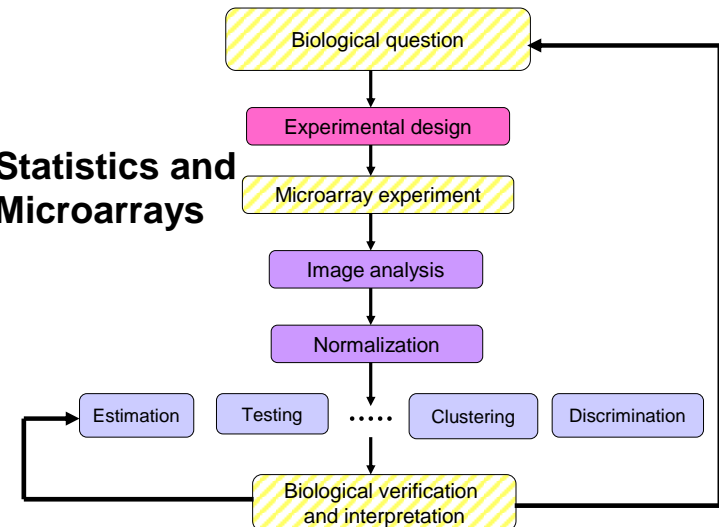
## Expression measures

- Most expression measures are based on differences of **PM-MM**.
- The intention is to correct for background and non-specific binding.
- E.g. MarrayArray Suite® (MAS) v. 4.0 uses Average Difference Intensity (ADI) or  $AvDiff = \text{average of PM-MM}$ .
- Problem: MM may also measure signal.
- More on this in lecture *Pre-processing in DNA microarray experiments*.

## What is the evidence?



## Statistics and Microarrays



## Statistical computing

### Everywhere ...

- for statistical design and analysis:
  - pre-processing, estimation, testing, clustering, prediction, etc.
- for integration with biological information resources (in house and external databases)
  - gene annotation (GenBank, LocusLink);
  - literature (PubMed);
  - graphical (pathways, chromosome maps).

## Integration of biological metadata

- Expression, sequence, structure, annotation, literature.
- Integration will depend on our using a common language and will rely on database methodology as well as statistical analyses.
- This area is largely unexplored.

## WWW resources

- **Complete guide to “microarraying”**  
<http://cmgm.stanford.edu/pbrown/mguide/>  
<http://www.microarrays.org>
  - Parts and assembly instructions for printer and scanner;
  - Protocols for sample prep;
  - Software;
  - Forum, etc.
- **cDNA microarray animation**  
<http://www.bio.davidson.edu/courses/genomics/chip/cchip.html>
- **Affymetrix**  
<http://www.affymetrix.com>

## Next ...

### *Pre-processing in DNA microarray experiments*

- cDNA microarrays
  - Image analysis;
  - Normalization.
- Affymetrix oligonucleotide chips
  - Image analysis;
  - Normalization;
  - Expression measures.