## Package 'awst'

October 16, 2024

Title Asymmetric Within-Sample Transformation

Version 1.12.0

License MIT + file LICENSE

Description We propose an Asymmetric Within-Sample Transformation (AWST) to regularize RNA-seq read counts and reduce the effect of noise on the classification of samples. AWST comprises two main steps: standardization and smoothing. These steps transform gene expression data to reduce the noise of the lowly expressed features, which suffer from background effects and low signal-to-noise ratio, and the influence of the highly expressed features, which may be the result of amplification bias and other experimental artifacts.

```
Encoding UTF-8
RoxygenNote 7.1.1
URL https://github.com/drisso/awst
BugReports https://github.com/drisso/awst/issues
Imports stats, methods, SummarizedExperiment
Suggests airway, ggplot2, testthat, EDASeq, knitr, BiocStyle,
      RefManageR, sessioninfo, rmarkdown
biocViews Normalization, GeneExpression, RNASeq, Software,
      Transcriptomics, Sequencing, SingleCell
VignetteBuilder knitr
git_url https://git.bioconductor.org/packages/awst
git_branch RELEASE_3_19
git_last_commit 8f7055e
git_last_commit_date 2024-04-30
Repository Bioconductor 3.19
Date/Publication 2024-10-16
Author Davide Risso [aut, cre, cph] (<a href="https://orcid.org/0000-0001-8508-5012">https://orcid.org/0000-0001-8508-5012</a>),
      Stefano Pagnotta [aut, cph] (<a href="https://orcid.org/0000-0002-8298-9777">https://orcid.org/0000-0002-8298-9777</a>)
Maintainer Davide Risso <risso.davide@gmail.com>
```

2 awst

### **Contents**

awst	Asymmetric Within-Sample Transformation	
Index		5
	awst	

#### **Description**

This function implements the asymmetric within-sample transformation described in Risso and Pagnotta (2019). The function includes two steps: a standardization step and a asymmetric winsorization step. See details.

#### Usage

```
## S4 method for signature 'matrix'
awst(x, poscount = FALSE, full_quantile = FALSE, sigma0 = 0.075, lambda = 13)

## S4 method for signature 'SummarizedExperiment'
awst(
    x,
    poscount = FALSE,
    full_quantile = FALSE,
    sigma0 = 0.075,
    lambda = 13,
    expr_values = "counts",
    name = "awst"
)
```

#### **Arguments**

X	a matrix of (possibly normalized) RNA-seq read counts or a 'SummarizedExperiment'.
poscount	a logical value indicating whether positive counts only should be used for the standardization step.
full_quantile	a logical value indicating whether the data have been normalized with the full-quantile normalization. In this case, computations can be sped up.
sigma0	a multiplicative constant to be applied to the smoothing function.
lambda	a parameter that controls the growth rate of the smoothing function.
expr_values	integer scalar or string indicating the assay that contains the matrix to use as input.
name	string specifying the name of the assay to be used to store the results of the transformation.

gene\_filter 3

#### **Details**

The standardization step is based on a log-normal distribution of the high-intensity genes. Optionally, only positive counts can be used in this step (this option is especially useful for single-cell data). The winsorization step is controlled by two parameters, sigma0 and lambda, which control the growth rate of the winsorization function.

#### Value

if 'x' is a matrix, it returns a matrix of transformed values, with genes in rows and samples in column. If 'x' is a 'SummarizedExperiment', it returns a 'SummarizedExperiment' with the transformed value in the 'name' slot.

#### Methods (by class)

- matrix: the input is a matrix of (possibly normalized) counts
- SummarizedExperiment: the input is a SummarizedExperiment with (possibly normalized) counts in one of its assays.

#### References

Risso and Pagnotta (2019). Within-sample standardization and asymmetric winsorization lead to accurate classification of RNA-seq expression profiles. Manuscript in preparation.

#### **Examples**

```
x \leftarrow matrix(data = rpois(100, lambda=5), ncol=10, nrow=10)
 awst(x)
```

gene\_filter

Gene filtering based on heterogeneity

#### **Description**

This function filters out genes that show a low heterogeneity, as measured by Shannon's entropy.

#### Usage

```
## $4 method for signature 'matrix'
gene_filter(
    x,
    from = min(x, na.rm = TRUE),
    to = max(x, na.rm = TRUE),
    nBins = 20,
    heterogeneity_threshold = 0.1
)
```

4 gene\_filter

```
## S4 method for signature 'SummarizedExperiment'
gene_filter(
    x,
    from = min(assay(x, awst_values), na.rm = TRUE),
    to = max(assay(x, awst_values), na.rm = TRUE),
    nBins = 20,
    heterogeneity_threshold = 0.1,
    awst_values = "awst"
)
```

#### **Arguments**

x a matrix of transformed gene expression counts (typically the results of awst).

from the minimum value from which to start binning data.

to the maximum value for the binning of the data.

nBins the number of bins.

heterogeneity\_threshold the trheshold used for the filtering.

awst\_values integer scalar or string indicating the assay that contains the awst-transformed values to use as input.

#### **Details**

Shannon's entropy is computed on the categorized data after AWST transformation. Those genes that show a lower entropy than the predefined threshold are deemed to carry too low information to be useful for the classification of the samples, and are hence removed.

#### Value

if 'x' is a matrix, it returns a filtered matrix. If 'x' is a 'SummarizedExperiment', it returns a filtered 'SummarizedExperiment'

#### Methods (by class)

- matrix: the input is a matrix of awst-transformed values.
- SummarizedExperiment: the input is a SummarizedExperiment with awst-transformed values in one of its assays.

#### References

Risso and Pagnotta (2019). Within-sample standardization and asymmetric winsorization lead to accurate classification of RNA-seq expression profiles. Manuscript in preparation.

#### **Examples**

```
set.seed(222)
x <- matrix(rpois(75, lambda=5), ncol=5, nrow=15)
a <- awst(x)
gene_filter(a)</pre>
```

# **Index**